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For Professional Use Only

AmpliSens® ***CCHFV*-FRT** PCR kit

Instruction Manual

AmpliSens®

EC REP

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1. INTENDED USE

AmpliSens® ***CCHFV*****-FRT** PCR kit is an *in vitro* nucleic acid amplification test for qualitative detection of *Crimean-Congo hemorrhagic fever virus (CCHFV)* RNA in clinical material (blood plasma and serum) and ticks using real-time hybridization-fluorescence detection of amplified products.

The results of PCR analysis are taken into account in complex diagnostics of disease.

2. PRINCIPLE OF PCR DETECTION

Crimean-Congo hemorrhagic fever virus detection includes:

1. RNA extraction from biological material sample;
2. Reverse transcription of RNA and amplification of *CCHFV* cDNA fragment with real-time hybridization-fluorescence detection.

CCHFV RNA detection by polymerase chain reaction (PCR) is based on the amplification of pathogen genome specific region using specific *CCHFV* primers. In the real-time PCR, the amplified product is detected with the use of fluorescent dyes. These dyes are linked to oligonucleotide probes, which bind specifically to the amplified product during thermocycling. The real-time monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

AmpliSens® ***CCHFV*****-FRT** PCR kit is a qualitative test that contains the Internal Control (Internal Control STI-87-rec (IC)). It must be used in the extraction procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition.

AmpliSens ® ***CCHFV*****-FRT** PCR kit uses “hot-start”, which greatly reduces the frequency of nonspecifically primed reactions. “Hot-start” is guaranteed by separation of nucleotides and Taq-polymerase by using a chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

3. CONTENT

AmpliSens® ***CCHFV*****-FRT** PCR kit is produced in 1 form:

AmpliSens® ***CCHFV*****-FRT** PCR kit variant FRT, **REF** R-V22-50-F(RG,iQ,Mx,Dt)-CE.

AmpliSens® *CCHFV*-FRT PCR kit variant FRT includes:

Reagent	*Description*	*Volume, ml*	*Amount*
RT-PCR-mix-1-FRT *CCHFV*	colorless clear liquid	0.6	1 tube
RT-PCR-mix-2-FEP/FRT	colorless clear liquid	0.3	1 tube
RT-G-mix-2	colorless clear liquid	0.015	1 tube
Polymerase (TaqF)	colorless clear liquid	0.03	1 tube
TM-Revertase (MMlv)	colorless clear liquid	0.015	1 tube
Positive Control cDNA *CCHFV* / STI (C+$_{CCHFV\ /\ STI}$)	colorless clear liquid	0.1	1 tube
RNA-buffer	colorless clear liquid	0.6	2 tubes
Negative Control (C–)*	colorless clear liquid	1.6	1 tube
Positive Control *CCHFV*-FL-rec**	colorless clear liquid	0.03	5 tubes
Internal Control STI-87-rec (IC)***	colorless clear liquid	0.12	5 tubes
tRNA 1 µg/µl	colorless clear liquid	0.06	5 tubes

* must be used in the extraction procedure as Negative Control of extraction (C–).

** must be used in the extraction procedure as Positive Control of extraction (PCE).

*** add 10 µl of Internal Control STI-87-rec during the RNA extraction procedure directly to the sample/lysis mixture (see RIBO-prep **REF** K2-9-Et-50-CE or RIBO-zol-B **REF** K2-3-50-CE protocol).

AmpliSens® *CCHFV*-FRT PCR kit is intended for 60 reactions (including controls).

4. ADDITIONAL REQUIREMENTS

- 0.15 M sodium chloride or phosphate buffer solution (PBS) (sodium chloride, 137mM; potassium chloride, 2.7 mM; sodium monophosphate, 10 mM; potassium diphosphate, 2 mM, pH=7.5±0.2).
- RNA/DNA extraction kit.
- Homogenizer (for ticks homogenization).
- Stainless steel beads (7 mm diameter).
- Disposable powder-free gloves and a laboratory coat.
- Pipettes (adjustable).
- Sterile RNase-free pipette tips with filters (up to 200 µl).
- Tube racks.
- Vortex mixer.

- Desktop centrifuge with a rotor for 2-ml reaction tubes.
- PCR box.
- Real-time instruments (for example, Rotor-Gene 3000, Rotor-Gene 6000 (Corbett Research, Australia), Rotor-Gene Q (QIAGEN, Germany), iCycler iQ5 (Bio-Rad, USA), or Mx3000P (Stratagene, USA) instrument).
- Disposable polypropylene tubes for PCR of 0.2- or 0.1-ml:
  a) 0.2-ml PCR tubes with optical transparent domed caps if a plate-type instrument is used;
  b) 0.2-ml PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Refrigerator with the temperature range from 2 to 8 °C.
- Deep-freezer with the temperature range from minus 24 to minus 16 °C.
- Reservoir for used tips.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (samples, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterward.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all samples and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in a biological cabinet in accordance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid samples and reagents contact with the skin, eyes and mucous membranes. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice immediately.
- Material Safety Data Sheets (MSDS) are available on request.

- Use of this product should be limited to personnel trained in the DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment, and reagents to the area where the previous step was performed.

Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

6. SAMPLING AND HANDLING

Obtaining samples of biological materials for PCR-analysis, transportation and storage is described in manufacturer's handbook [1]. It is recommended that this handbook is read before starting work.

AmpliSens® ***CCHFV*-FRT** PCR kit is intended for analysis of the RNA extracted with RNA/DNA extraction kit from clinical material (blood plasma and serum) and ticks:

6.1. *Blood plasma and blood serum.* Take samples of peripheral blood after overnight fasting into the tube with 6% EDTA solution at a proportion of 1:20. Turn over the closed tube with a blood sample several times. Centrifuge the tube with a blood sample at 1600 g for 20 min to obtain a plasma sample. Collect a serum sample in accordance with the standard procedure. Take 100 µl of clinical material for the analysis.

6.2. *Ticks.* Before tick pretreatment, pools of ticks should be formed. Each pool can contain 5-7 non-sated ticks, 2-3 ticks of semi-sated ticks, or 1 fully sated one. Use sterile porcelain mortars and sterile pestles for ticks suspension preparation. If an automatic homogenizer TissueLyser LT is used, the following parameters are set for the ticks of *Hyalomma* genus: diameter of beads – 7 mm; frequency – 50 Hz/s; homogenization time – 12-15 min; buffer volume – 700 µl (non-sated ticks), 1,000-1,500 µl (sated ticks and pools). Prior to homogenization, sated ticks should be pierced with a sterile disposable needle to let blood out. Oily ticks should be washed with 70 % ethanol solution. Homogenize the ticks in 700 µl (if a sample consists of 1 non-sated tick) or 1-1.5 ml (for a tick pool or a sated tick) of 0.15 M sodium chloride or PBS buffer. Add the solution by small portions. Centrifuge the obtained suspension at 10,000 g for 1 min. Take 50 µl of the supernatant for RNA extraction with RIBO-prep reagent kit. RNA from sated ticks should be extracted with RIBO-zol-B reagent kit. In this case, 100 µl of the supernatant is used.

Before analysis, the biological material can be stored at 2–8 °C for 1 day, at ≤ –16 °C for

1 week. Ticks can be stored alive (up to 1 month) or at ≤ –16 °C for 1 week and then at ≤ – 70 °C.

7. WORKING CONDITIONS

AmpliSens® ***CCHFV*****-FRT** PCR kit should be used at 18–25 °C.

8. PROTOCOL

8.1. RNA extraction

It is recommended that the following nucleic acid extraction kits are used:

- RIBO-prep, **REF** K2-9-Et-50-CE – for RNA extraction from blood plasma and serum or suspension of non-sated or semi-sated ticks.
- RIBO-zol-B **REF** K2-3-50-CE – for RNA extraction from suspension of sated ticks.

8.1.1 RNA extraction with the use of RIBO-prep reagent kit:

Extract the RNA according to the manufacturer's protocol.

Volume of a **tick suspension** sample is **50 µl**.
Volume of a **blood plasma/serum** sample is **100 µl**.

To the tube intended for the Positive Control of extraction (PCE) add
10 µl of Internal Control STI-87-rec,
300 µl of Solution for Lysis, and
10 µl of Positive Control *CCHFV*-FL-rec

To the tube intended for the Negative Control of extraction (C–) add
10 µl of Internal Control STI-87-rec and
300 µl of Solution for Lysis

8.1.2 RNA extraction with the use of RIBO-zol-B reagent kit:

1. Take the required number of 1.5-ml screw cap tubes (including the Positive and Negative Controls of extraction). Add **10 µl** of **Internal Control STI-87-rec** to each tube intended for extraction from test material then add **300 µl** of **Solution D**. Label the tubes.
2. Add **100 µl** of **tick suspension** to the tubes with Internal Control STI-87-rec and Solution D.
3. To the tube intended for the Positive Control of extraction (PCE) add:

   10 µl of Internal Control STI-87-rec,
   300 µl of Solution D,
   80 µl of Negative Control, and
   10 µl of Positive Control *CCHFV*-FL-rec.

4. To the tube intended for the Negative Control of extraction (C–) add:

   10 µl of Internal Control STI-87-rec,
   300 µl of Solution D, and
   90 µl of **Negative Control**.
5. Thoroughly vortex the tubes and then incubate at **56 °C for 5 min**. Centrifuge the tubes to remove drops from the tube walls.
6. Add **30 µl** of **Solution E**. Vortex the tubes then centrifuge to remove drops from the tube walls.
7. Add **300 µl** of **Solution A**. Vortex the tubes then centrifuge to remove drops from the tube walls.
8. Add **100 µl** of **Solution B**. Vortex the tubes for 1–2 min (solution should become milky).
9. Place the tubes in a refrigerator at 2– 4 °C for 10 min.
10. Centrifuge the tubes at 10,000 g for 10 min. Solution should separate in 2 phases: the bottom phase containing proteins and DNA and the top aqueous phase containing RNA.
11. Take new 1.5-ml tubes and add **300 µl** of **Solution C**. Label the tubes. Add **10 µl** of **tRNA 1 µg/µl** to the tubes intended for C– and PCE.
12. Carefully remove the top phase (about 400 µl) using tips with aerosol barrier and transfer it into the tube with Solution C. Vortex the tubes and incubate in a deep-freezer at ≤ –16 °C for 1 h.
13. Centrifuge the tubes at 10,000 g for 10 min. Remove the supernatant using a 1-ml tip (do not disturb the pellet).
14. Dilute the pellet in **100 µl** of **Solution D**, add **100 µl** of **Solution C**, and vortex. Incubate in a deep-freezer at ≤ –16 °C for 1 h.
15. Centrifuge the tubes at 10,000 g for 10 min. Remove the supernatant using a 1-ml tip (do not disturb the pellet).
16. Wash the pellet in **800 µl** of **Washing Solution 3** cooled at 2–8 °C by vortexing. Then centrifuge the tubes at 10,000 g for 10 min. Remove the supernatant using a 1 ml tip (do not disturb the pellet).
17. Add **150 µl** of cooled **Washing Solution 3**. Centrifuge the tubes at 10,000 g for 10 min. Remove the supernatant using a 200-µl tip (do not disturb the pellet).
18. Incubate the tubes at 56 °C for 5 min to dry the pellet. Make sure that the tubes are open.
19. Add **50 µl** of **RNA-eluent** into the tubes. Dilute RNA pellet by vortexing. Bring RNA-eluent volume to 100 µl in case of high viscosity of the solution. Incubate the tubes

for 5-7 min.

20. Centrifuge the tubes at 10,000 g for 2 min. The supernatant contains purified RNA ready for reverse transcription and PCR.

RNA solution is to be stored at ≤ –68 °C.

8.2. Preparing the tubes for reverse transcription and PCR

The total reaction volume is **25 µl**, the volume of the RNA sample is **10 µl**.

1. Before starting work, thaw and thoroughly vortex all reagents of the kit. Make sure that there are no drops on the caps of the tubes.
2. Take the required number of PCR tubes for amplification of clinical and control samples (including two controls of extraction, PCE and C–, and two controls of reverse transcription and amplification, C+, and NCA). The type of tubes depends on the real-time PCR instrument used for the analysis.
3. Prepare the reaction mixture for the required number of reactions. To do this, mix in a new tube RT-PCR-mix-1-FRT *CCHFV*, RT-PCR-mix-2-FEP/FRT, polymerase (TaqF), TM-Revertase (MMlv), and RT-G-mix-2 in the following proportion (the calculation is specified for one reaction):
   - **10 µl of RT-PCR-mix-1-FRT *CCHFV*,**
   - **5 µl of RT-PCR-mix-2-FEP/FRT**,
   - **0.5 µl of polymerase (TaqF)**,
   - **0.25 µl of RT-G-mix-2**,
   - **0.25 µl of TM-Revertase (MMlv)**.
4. Add **15 µl** of the prepared reaction mixture to each PCR tube.

Do not store the prepared mixture.

5. Add **10 µl** of **RNA samples** extracted from the clinical and control samples to each PCR tube. Carefully mix by pipetting.
6. Run the **control reactions**:

NCA -Add **10 µl** of **RNA-buffer** to the tube labeled NCA (Negative Control of Amplification)

C+ –Add **10 µl** of **Positive Control cDNA *CCHFV* / STI (C+$_{CCHFV\ /\ STI}$)** to the tube labeled C+ (Positive Control of Amplification)

Samples should be amplified immediately after mixing the reaction mixture with RNA samples and control samples.

8.2.2 Reverse transcription and amplification

1. Create a temperature profile on your instrument as follows:

Table 1

	Rotor-type instruments[1]			Plate-type instruments[2]		
Cycle	Temperature°C	Time	Cycle repeats	Temperature °C	Time	Cycle repeats
1	**50**	30 min	1	**50**	30 min	1
2	**95**	15 min	1	**95**	15 min	1
3	**95**	10 s	5	**95**	10 s	5
	54	25 s		**54**	30 s	
	72	15 s		**72**	15 s	
4	**95**	10 s	45	**95**	10 s	45
	50	25 s Fluorescence acquiring		**50**	35 s Fluorescence acquiring	
	72	15 s		**72**	15 s	

Fluorescent signal is detected in the channels for the FAM and JOE fluorophores.

2. Adjust the fluorescence channel sensitivity according to the *Important Product Information Bulletin* and Guidelines [2].
3. Insert the tubes into the reaction module of the device. **Well No. 1 should be loaded with a test tube.**
4. Run the amplification program with fluorescence detection.
5. Analyze results after the amplification program is completed.

9. DATA ANALYSIS

The result is interpreted by software of the used real-time instrument. The curves of fluorescence signal accumulation are analyzed in two channels:

– The signal of the IC cDNA amplification product is detected in the channel for the FAM fluorophore.
– The signal of the *CCHFV* cDNA amplification product is detected in the channel for the JOE fluorophore.

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a *Ct* value of the cDNA sample in the corresponding column of the results grid.

[1] For example, Rotor-Gene 3000/6000 (Corbett Research, Australia), Rotor-Gene Q (Qiagen, Germany).
[2] For example, iCycler iQ5 (Bio-Rad, USA), Mx3000P (Stratagene, USA).

Principle of interpretation is the following:

- *CCHFV* cDNA is **detected** if *Ct* value determined in the channel for the JOE fluorophore does not exceed the specified boundary *Ct* value. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- *CCHFV* cDNA is **not detected** if *Ct* value determined in the channel for the FAM fluorophore does not exceed the specified boundary *Ct* value, whereas *Ct* value in the channel for the JOE fluorophore is not determined or exceeds the specified boundary value.
- the result is **invalid** if *Ct* value in the channel for the JOE fluorophore is not determined (absent) and *Ct* value in the channel for the FAM fluorophore is not determined or exceeds the specified boundary value. In this case, PCR analysis of the required sample should be repeated beginning with the extraction.

Boundary *Ct* values are specified in the *Important Product Information Bulletin* enclosed in the PCR kit. See also Guidelines [2]

The result of the analysis is considered reliable only if the results obtained for the Positive and Negative Controls of Amplification as well as for the Positive and Negative Controls of Extraction are correct (see table 2).

Table 2

Results for controls

Control	**Stage for control**	***Ct* in channel**	
		FAM	**JOE**
C–	RNA extraction	**<**boundary value	Absent
PCE	RNA extraction	**<**boundary value	**<**boundary value
NCA	Amplification	Absent	Absent
C+	Amplification	**<**boundary value	**<**boundary value

10. TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

1. If the *Ct* value determined for the Positive Control of Amplification (C+) in the channel for the **JOE** fluorophore is absent or exceeds the boundary value, amplification of all samples in which *CCHFV* cDNA was not detected should be repeated once again.
2. If the *Ct* value determined for the Positive Control of extraction (PCE) in the channel for the **JOE** fluorophore is absent or exceeds the specified boundary value, extraction of all samples in which *CCHFV* cDNA was not detected should be repeated once again.
3. If the *Ct* value is determined for Negative Control of extraction (C–) in the channel for the **JOE** fluorophore PCR analysis should be repeated for all samples in which cDNA was

detected in the channel for the **JOE** fluorophore.

4. If the *Ct* value is detected for Negative Control of amplification (NCA) in the channels for the **FAM** and **JOE** fluorophores, amplification of all samples in which cDNA was detected in the channel for the **JOE** fluorophore should be repeated once again accompanied with amplification of NCA sample in three replicates.

If you have any further questions or if you encounter problems, please contact our Authorized representative in the European Community.

11. TRANSPORTATION

AmpliSens® ***CCHFV*-FRT** PCR kit should be transported at 2–8 ºC for no longer than 5 days.

12. STABILITY AND STORAGE

All components of the **AmpliSens®** ***CCHFV*-FRT** PCR kit are to be stored at 2–8 °C when not in use (except for RT-G-mix-2, RT-PCR-mix-1-FRT *CCHFV,* RT-PCR-mix-2-FEP/FRT, polymerase (TaqF), TM-Revertase (MMlv), and tRNA 1µg/µl). All components of the **AmpliSens®** ***CCHFV*-FRT** PCR kit are stable until the expiration date on the label. The shelf life of reagents before and after the first use is the same, unless otherwise stated.

RT-G-mix-2, RT-PCR-mix-1-FRT *CCHFV,* RT-PCR-mix-2-FEP/FRT, polymerase (TaqF), TM-Revertase (MMlv), and tRNA 1µg/µl are to be stored at temperature from minus 24 to minus 16 °C when not in use.

RT-PCR-mix-1-FRT *CCHFV* is to be kept away from light.

13. SPECIFICATIONS

13.1. Sensitivity

The analytical sensitivity of **AmpliSens®** ***CCHFV*-FRT** PCR kit is specified in the table below.

Test material (sample volume)	RNA/DNA extraction kit	Analytical sensitivity, copies/ml	Pretreatment
Blood serum (100 µl)	RIBO-prep	$5x10^3$	The claimed sensitivity is achieved only when the material pretreatment is carried out in accordance with chapter *Sampling and Handling* and the recommended volume of test sample is used
H.marginatum tick pools (50 µl)			
H.marginatum ticks pools (100 µl)	RIBO-zol-B	$5x10^3$	

13.2. Specificity

The analytical specificity of **AmpliSens® *CCHFV*-FRT** PCR kit is ensured by selection of specific primers and probes and stringent reaction conditions. The primers and probes were checked for possible homologies to all sequences deposited in gene banks by sequence comparison analysis. Analytical specificity was examined with the use of the following microorganisms:

- *flaviviruses* (*West Nile fever virus*, *Omsk hemorrhagic fever virus*);
- *herpesviruses* (types I and II, *cytomegalovirus*; *Epstein-Barr virus*, *Varicella-Zoster virus*, type IV), *enteroviruses* (*ECHO*, *Coxsackie*);
- *rickettsiae* of the spotted fever group (*Rickettsia conorii* ssp*. caspia*, *Coxiella burnetii);*
- *orthobunyaviruses (Tyaginya virus, Batai virus);*
- *hantaviruses* (*Puumala virus, Dobrava virus*);
- *thogotoviruses (Batken virus).*

False positive results for the above-mentioned organisms and viruses as well as human DNA and tick DNA were not detected.

The clinical specificity of **AmpliSens® *CCHFV*-FRT** PCR kit was confirmed in laboratory clinical trials.

14. REFERENCES

1. Handbook “Sampling, Transportation, and Storage of Clinical Material for PCR diagnostics”, developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology” of Federal Service for Surveillance on Consumers’ Rights Protection and Human Well-Being, Moscow, 2010.
2. Guidelines to the **AmpliSens® *CCHFV*-FRT** PCR kit for qualitative detection of *Crimean-Congo hemorrhagic fever virus (CCHFV)* RNA in clinical material (blood plasma and serum) and ticks by real-time hybridization-fluorescence detection of amplified products developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology”.

15. QUALITY CONTROL

In compliance with the Federal Budget Institution of Science “Central Research Institute for Epidemiology” ISO 13485-Certified Quality Management System, each lot of **AmpliSens® *CCHFV*-FRT** PCR kit has been tested against predetermined specifications to ensure consistent product quality.

16. KEY TO SYMBOLS USED

REF	Catalogue number		Caution
LOT	Batch code	Σ	Sufficient for
IVD	*In vitro* diagnostic medical device		Expiration Date
VER	Version		Consult instructions for use
	Temperature limitation		Keep away from sunlight
	Manufacturer	**NCA**	Negative control of amplification
	Date of manufacture	**C–**	Negative control of extraction
CCHFV	*Crimean-Congo hemorrhagic fever virus*	**C+**	Positive control of amplification
EC REP	Authorised representative in the European Community	**IC**	Internal control
		PCE	Positive Control of extraction